

NEW METHODS TO IMPROVE THE BIOECONOMIC AND ECOECONOMIC IMPACT BY PLANT BIOTECHNOLOGY

Adriana Petruș-Vancea*¹

¹University of Oradea and postdoctoral researcher at "Vasile Goldis" West University of Arad, Romania

ABSTRACT.

Sugar beet (*Saccharomices cerevisiae* var. *Saccharifera*) is a species of economic and food interest, which, in terms of clonally multiplication, is liable to hyperhydricity. In these studies was identified a method that prevents the hyperhydricity process installation, and at the same time, it increased *in vitro* plant biomass, using honey, as a replacement for sucrose, and deuterium depleted water (DDW), as a replacement for distilled water (DW), either culture in single-sided, or in the double layer system. Murashige-Skoog mineral culture media, solid, free of growth regulators, prepared with DW or DDW, with the supernatant consist in diluted honey either DW or in DDW were most beneficial in stimulating both the rhyso-genesis and the caulogenesis process. The increase of plant biomass in the vitroculture and eliminating waste generated by hyperhydricity has a positive economic impact for producers in this branch.

Keywords: deuterium, honey, double system layer

INTRODUCTION

Plant biotechnology is a domain with a great practical value, it represent the solutions to solve the major social problems of humankind such as providing food and energy, health and environmental protection, in the current context of the bioeconomy, ecoeconomy and ecosanogenesis. Modern and traditional cultures is essential for further supporting food security and new policies (Bogdan et al., 2010; Ipate et al., 2010) in relation with biodiversity conservation and climate change responses such as adaptation and mitigation (Antofie et al., 2010 b).

Identification of new methods in plant biotechnology with applications in food and not only has an important step, predicts the ecoeconomic and bioeconomic impact of food safety and security in perspective the increased consumption of food and feed during 2030-2100. The dynamic of food's world population projection to 2050 year and consumption dynamic of vegetables in terms of respect international standards of food safety and security (Bogdan et al., 2010). In this framework, the food safety and security must be correlated with the respect of known principles of Hazard Analysis and Critical Control Points, based on actual international standards from ISO 9001-9002 series (Quality Management System), ISO 14001: 2004 (Environment Management), ISO 22000 (Food Safety).

Sugar beet (*Beta vulgaris* var. *Saccharifera*) has economic interest (Atanasov, 1986) for sugar production, required for both food and ethanol obtain, and which, for conservation [among other techniques developed *in situ* (Antofie, 2010; Antofie et al., 2010 a; Purcărea and Borbely, 2011)] and for amelioration, is cloned by micropropagation procedures. Beet multiplications was achieved through various types of inoculs, namely the apex in the early seed-lobe, the buds from colet or from shoots (Sand and Cachiță, 1999; Sand and Cachiță, 2000),

phytoinocula placed on different types of culture media with or without added growth regulators or in which sucrose was partially replaced, successful, with starch (Constantinovici, 1997; Constantinovici and Cachiță, 1999). In such practices, the hyperhydricity appearance on vitroculture is quite common for this species, which lowers production yield for the field of plant breeding, plant which are indispensable in rhyso-mania resistant genotypes selecting and their cloning. At phytoinocula, hyperhydricity is considered a complex neoplastic disease process, reflected not only by an abnormal accumulation of water in tissue, but also by the anatomical changes appearance and deep morpho-physiological organs affected by this syndrome, which is translated and a tissue hypertrophy and cells hyperplasia (Cachiță et al., 2008 a, 2009).

In some experiment, Cachiță and collaborators (2008 a, 2009) has highlighted the profound transformation suffered by sugar beet hyperhydric vitroplantlets, both in altering terms of the chloroplasts structure and in a fast decrease of the assimilating pigments contents, especially in chlorophyll *a*. In hyperhydric vitroplantlets leaflets foliar mesophyll cells this authors identified either chloroplasts flatten or volume increased, but with a content being disorganization; they were scattered in vacuolar juice mixed with cytoplasm (myxoplasm), a mixture resulting because the tonoplast disintegration. The hyperhydric leaflets presented a low content of assimilating pigments, much lower than that determined in sugar beet greenhouse plantlet leaflets, and lower than that recorded in leaflets of „in vitro” nonhyperhydric plantlets, which was treated with DDW. DDW can prevent and annihilate hyperhydricity (Cachiță et al., 2008 b and Cachiță et al., 2010).

In this research we intend to stimulate growth of sugar beet plant biomass, without hyperhydricity by replacing sucrose as carbon source, with acacia honey,

* **Correspondence:** Adriana Petrus-Vancea, University of Oradea, Faculty of Science, Biology Department, no 1, Universitatii St., Oradea, Bihor, Romania, 410082, E-mail: adrianaivan@yahoo.com

either single-sided, or in the double layer system. Present research is an answer at the question that arises more and more, in present: whether and under what conditions the estimated future food demand can be met and how food security can be achieved.

MATERIAL AND METHODS

The phytoinocules consisted in sugar beet (*Beta vulgaris* var. *Saccharifera*) uninodal, apical minicuttings,

Single-layer system

- V₀ - BM-MS solid prepared with DW and sucrose (control);
- V₁ - BM -MS prepared with DDW and sucrose;
- V₂ - BM -MS prepared with DW and honey;
- V₃ - BM -MS prepared with DDW and honey.

Double-layer system

- V₄ - BM -MS prepared with DW and sucrose + supernatant DDW;
 - V₅ - BM -MS prepared with DW and sucrose + supernatant honey, mixed with DW;
 - V₆ - BM -MS prepared with DW and sucrose + supernatant honey, mixed with DDW.
-

We were taken into account that to each experimental variant, the carbohydrate content (regardless of its nature) to a total of 30 g/l, both for single layer and double layer cultures (Cachiță, 1987).

Vitrocultures vessels consisted of glass jars, 7 cm height and 4 cm diameter, each one with 20 ml of solid medium (first layer). The supernatant was 5 ml.

Incubation and growth was realised in growth chamber at 22 ° C ± 2 ° C, illuminated with white fluorescent tubes, 16h day length photoperiod of 24 h day light and 1700 lx intensity.

The biometrisation of plantlets growth indices were realised at 90 days after inoculation and the dry weight were weighed after maintenance in aluminium foil, in oven at 115 °C, for 3 days.

The dates obtained from measurements were interpreted statistically by analysis of variance, also the statistical significance was determined using *one sample t test* of statistical SPSS for Windows.

RESULTS AND DISCUSSIONS

At 90 days after inoculation, the sugar beet vitroplantlets rhylogenesis was decreased to control lot (V₁) and those in single-layer, in which the DDW was replaced with DW (Table 1). Moreover, the phytoinocula submerged in the supernatant consisting of DDW (V₅) showed *in vitro* no regeneration processes.

The same rhylogenesis inhibition effect exerted by DDW was reported by Petruș-Vancea and Cachiță

which provided from aseptically germinated seeds, grown on basic medium (BM) Murashige-Skoog (MS) (1962) ½, solid, without growth regulators.

The culture medium used in these experiments was MS (1962), modified by us; the solidification was achieved by 7 g/l agar-agar; culture medium pH was adjusted to a value of 5.7 before its autoclaving.

The culture medium preparation was performed according to the following test:

(2008 and 2009) in the case of chrysanthemum or African violets plantlets base watering with this type of water, throughout the period of acclimatization to the septic medium. Also, rhylogenesis to the *Tradescantia* cuttings, rooted in natural living conditions in perlite, was inhibited by watering them with DDW, compared with watering with DW (Petruș and Cachiță, 2008). At the *Cymbidium hybridum* and *Petunia* cultures, placed on MS medium (1962), in single-layer system, solid, without growth regulators, DDW was a timing effect of growth, with an important role in *in vitro* preserving cultures (Petruș et al., 2004). Also mixture of honey and DDW - but as supernatant - was beneficial in the caulogenesis stimulation, the longest stemlets, an average of 2.11 cm, and a large number of leaflets, few of them being necrotic were registered at vitroplantlets cultivated on experimental variant denoted V₆. Mediums containing honey and DDW generated vitroplantlets with fresh weight, respectively dry weight increased, on medium in which the mixture was consist in solid layer, on the base system (V₃), the rhylogenesis was stimulated, and at the regenerated vitroplantlets in the supernatant consisting of those two component (V₆), the caulogenesis being intensified (Table 1), while separate use of two natural compounds generated even total inhibition of *in vitro* regeneration, as in DDW case which was used individually as supernatant (V₄).

Table 1.

Comparative analysis of average values of sugar beet (*Beta vulgaris* var. *Saccharifera*) vitroplantlets growth indices, at 90 days after inoculation, as follows: V₀ - BM-MS solid prepared with DW and sucrose (control); V₁ - BM-MS prepared with DDW and sucrose; V₂ - BM-MS prepared with DW and honey; V₃ - BM-MS prepared with DDW and honey; V₄ - BM-MS, prepared with DW and sucrose + supernatant DDW; V₅ - BM-MS, prepared with DW and sucrose + supernatant honey, mixed with DW; V₆ - BM-MS, prepared with DW and sucrose + supernatant honeyl, mixed with DDW.

Biometri- sation	V ₀		V ₁		V ₂		V ₃		V ₄		V ₅		V ₆	
	Mean± st.dev	Sig	Mean± st.dev	Sig	Mean± st.dev	Sig	Mean± st.dev	Sig	Mean± st.dev	Sig	Mean± st.dev	Sig	Mean± st.dev	Sig
Rootlet no.	2.85± 2.27	***	2.56± 1.82	***	5.38± 1.12	***	9.00± 1.61	***	-	-	8.50± 1.58	***	6.44± 3.24	***
Rootlet L. (cm)	1.96± 1.05	***	1.19± 1.29	**	0.73± 0.44	***	1.55± 0.52	***	-	-	1.75± 0.86	***	1.67± 1.00	***
Stem L. (cm)	1.46± 0.14	***	1.19± 0.23	***	1.23± 0.26	***	1.95± 0.15	***	-	-	1.95± 0.28	***	2.11± 0.49	***
Leaflets total no.	9.62± 1.80	***	7.50± 1.79	***	8.08± 1.26	***	9.27± 1.01	***	-	-	9.20± 1.03	***	9.33± 1.00	***
Necrosed leaflets no.	3.54± 2.22	***	2.50± 1.79	***	1.31± 0.48	***	1.64± 0.67	***	-	-	1.80± 0.92	***	1.56± 1.01	**
Petiole L.	3.19± 0.75	***	3.84± 0.93	***	4.15± 0.90	***	6.45± 0.52	***	-	-	5.50± 1.58	***	5.89± 1.05	***
Ramifica- tion no.	0.23± 0.44	*	0.06± 0.25	ns	0.31± 0.48	*	0.00± 0.00	***	-	-	0.10± 0.32	ns	0.00± 0.00	***
Fresh w.(g)	0.13± 0.01	***	0.24 ± 0.01	***	0.35 ± 0.01	***	0.96 ± 0.01	***	-	-	0.61± 0.20	***	0.93± 0.03	***
Dry w. (g)	0.02± 0.01	***	0.04 ± 0.01	***	0.04± 0.03	***	0.09 ± 0.01	***	-	-	0.06± 0.04	***	0.098± 0.01	***

Note: BM-MS – basal medium Murashige-Skoog (1962); DW – distilled water; DDW-deuterium depleted water; sig. – significance [ns – non significant (p > 0.1), ** significant (p < 0.05), *** very significant (p < 0.01)]; st.dev. – standard deviation; no – number; L – length; w – weight.

At *Cymbidium* protocorms (Vancea and Pătru, 2002) or at chrysanthemums and African violets plantlets (Petruş-Vancea et al., 2008) have been reported light inhibitions of *in vitro* organogenesis, being cultured on medium in which sucrose was replaced with acacia honey 20g/l, but when transferring the plantlets, regardless of species, living in septic environment, the survival of lot provided from honey was superior to that whose carbon source was sucrose.

In this research, the optimal culture medium for beet proved to be that which was prepared with honey, in which DW was replaced with DDW (V₃). Although the double layer media have generated intense growth of sugar beet vitroplantlets, as sparrow grass (personal dates), compared with this specie (which grows as a shrub), double layer system applied to beet has shown some mechanical drawbacks. Thus, the supernatant exerted a beneficial physiological effect on vitroplantlets growth, but, because the sugar beet leaflets, having long stalks, the plant arrived in the supernatant and it was overturned. Certainly, finding a support for plant (like luffa lignoskeleton described by Petruş - Vancea et al. 2004) as a solution to ensure the aerial part of vitroplantlets

and thus the optimization of micropropagation on double layer system from physically point of view.

CONCLUSIONS

Economic implications of the processes identified in this research are important, because we managed to remove two of the most delicate issues of sugar beet *in vitro* culture, namely hyperhydricity and quantity of plant biomass, due to successful combination of DDW and honey. Combining honey with DDW we managed to achieve two major objectives in the micropropagation of sugar beet, namely increasing biomass on vitrocultures and hyperhydricity prevention, thus eliminating loss of production and reducing the costs of obtaining healthy plant material, the bioeconomical and ecoeconomical impact being certainly.

ACKNOWLEDGEMENTS

This work was cofinanced from the European Social Fund through Sectoral Operational Programme Human Resources Development 2007-2013, project number POSDRU/89/1.5/S/63258 "Postdoctoral school for zootechnical biodiversity and food biotechnology based on the eco-economy and the bio-economy required by

ecosanogenesis”, coordinated by the National Institute of Economic Research ”Costin C. Kirițescu”, Romania, in collaboration with ”Lucian Blaga” University of Sibiu, ”Vasile Goldiș” West University Arad, S.C. **GNIR Holding S.A. and S.C. SIAT S.A.**

REFERENCES

- Antofie MM, Questionnaire for investigating on farm conservation status of old crops varieties - developing method. *Analele Universității din Oradea Fascicula: Ecotoxicologie, Zootehnie și Tehnologii de Industrie Alimentară*, IX, 1019 - 1026, 2010.
- Antofie MM, Pop, MR, Sand C, Ciotea G, Iagrăru P, Data sheet model for developing a red list regarding crop landraces in Romania. *Annals. Food Science and Technology*, 11(1), 45-49, 2010 a.
- Antofie MM, Constantinovici D, Pop MR, Iagrăru P, Sand C, Ciotea G, Theoretical methodology for assessing the status of conservation of crop landraces in Romania. *Analele Universității din Oradea – Fascicula Biologie*, XVII (2), 313-317, 2010 b.
- Atanassov A, Sugar beet (*Beta vulgaris* L.). In: Bajaj YPS (ed.), *Biotechnology in Agriculture and Forestry*, Vol. 2: Crops I, Springer - Verlag, Berlin, Heidelberg, 462 – 470, 1986.
- Bogdan AT, Ipate I, Bara V, Diaconescu D, Purcarea C, Strateanu AG, Ecoeconomic and bioeconomic impact of food safety and security in perspective increased consumption of food and feed during 2030-2100. *Analele Universității din Oradea Fascicula: Ecotoxicologie, Zootehnie și Tehnologii de Industrie Alimentară*, IX, 1044 – 1057, 2010.
- Cachiță CD, Metode *in vitro* la plantele de cultură. Baze teoretice și practice. Editura Ceres București, 1987.
- Cachiță CD, Petruș - Vancea A, Crăciun C, Issues regarding the chloroplast ultrastructure and assimilating pigments content in normal or hyperhydric sugar beet (*Beta vulgaris* L. var. *Saccharifera*) vitroplantlet leaflets. *Studia Univ. Vasile Goldiș, Seria Șt. Vieții*, 19(2), 287 – 294, 2009.
- Cachita CD, Petrus-Vancea A, Radovet D, Proceduri de prevenire sau de anihilare a hiperhidriei la fitoinoculi, prin utilizarea apei sărăcite în deuterium. Număr Patent RO 125649-A2, din/30.08.2010, International Patent Classification A01N-02500; C01B005/00, 2010.
- Cachiță CD, Petruș CM, Petruș-Vancea A, Crăciun C, Fenomenul de hiperhidrie manifest la nivelul vitroculturilor de sfeclă de zahăr (*Beta vulgaris* L. var. *Saccharifera*). II. Aspecte ultrastructurale identificate în celulele țesuturilor limburilor foliare ale plantulelor normale sau ale vitroplantulelor hiperhidrice. *Analele SNBC*, vol. XIII, cap. II. Date morfofiziologice și biochimice, 113-127, 2008 a.
- Cachiță CD, Petruș CM, Petruș - Vancea A, Crăciun C, Hyperhydricity phenomenon developed at the level of sugar beet (*Beta vulgaris* L. var. *Saccharifera*) vitrocultures. IV. Deuterium depleted water effects in hyperhydricity annihilation of vitroleaflet cells, aspects viewed to optic and transmission electronic microscope. *Studia Univ. Vasile Goldiș, Seria Șt. Vieții*, 18(2), 25-34, 2008 b.
- Constantinovici D, Studii privind efectul acțiunii „in vitro” a mediilor de cultură, cu diferite balanțe hormonale, asupra unor categorii de explante la *Beta vulgaris* L.. *Cercet. Agr. în Mold.*, 110, 71 – 79, 1997.
- Constantinovici D, Cachiță CD, Multiplicarea la sfecla de zahăr (*Beta vulgaris*) pe medii de cultură solidificate cu agar-agar substituit, parțial, cu amidon. In: Cachiță CD, Ardelean A, Crăciun C (eds.), *Lucrările reunite ale celui de al VII-lea și al VIII-lea Simpozion de Culturi de Țesuturi și Celule Vegetale, Arad 1997 și Buziaș 1998*, intitulat ”Culturi in vitro la cormofite”, Editura Risoprint Cluj-Napoca, 162 – 173, 1999.
- Ipate I, Bogdan AT, Paraschivescu M, Sandu M, Ivana S, Ipate N, Strateanu AG, Toba G, Enache M, Use rare breed for genuine foods in Romanian rural tourism and possibility of traceability the traditional products. *Bulletin UASVM Animal Science and Biotechnologies*, 67(1-2), 225-230, 2010.
- Murashige T, Skoog F, A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol. Plant* 15, 473 – 477, 1962.
- Petruș CM, Cachiță CD, 2008, Foliar and radicular sprinkling of *Tradescantia* cuttings, with different types of water, and their effect to organogenesis and the epidermal formations of foliar limbs. *Analele Univ. Oradea, Fasc. Biol.*, XV, 73-78, 1962.
- Petruș CM, Petruș - Vancea A, Cachiță CD, Micropropagarea la *Cymbidium* și *Petunia* pe medii de cultură preparate cu apă sărăcită în deuteriu. In: Cachiță CD, Ardelean A (eds.), *Lucrările celui de-al XII -lea Simpozion Național de Culturi de Țesuturi și Celule, Vegetale, Jibou*, intitulat ”Fiziopatologia celulei vegetale în regim de vitrokultură”, Editura Daya Satu – Mare, 185 – 192, 2004.
- Petruș - Vancea A, Cachiță CD, Regards on *Chrysanthemum* (*Chrysanthemum morifolium* Ramat var. *Lamet*), leafs epidermis which were sprinkle, during “ex vitro” acclimatization period, with Pi water and deuterium depleted water. *Studia Univ. Vasile Goldiș, Seria Șt. Vieții* 18, 81 – 86, 2008.

- Petruș – Vancea A, Cachiță CD, The effect of Pi or deuterium depleted water on foliar stomata of *Saintpaulia ionantha* L. exvitroplantlets sprinkled foliar or radicular. *Studia Univ. Vasile Goldiș, Seria Șt. Vieții*, 19(2), 309 – 312, 2009.
- Petruș - Vancea A, Cachiță CD, Blidar CF, Biogelul și *lignoskeletonul* de *lufă*, noi proceduri de vitrocultură. In: Cachiță CD, Ardelean A (eds.), *Lucrările celui de-al XII -lea Simpozion Național de Culturi de Țesuturi și Celule Vegetale*, Jibou, intitulat „Fiziopatologia celulei vegetale în regim de vitrocultură”, Editura Daya Satu – Mare, 171 – 177, 2004.
- Petruș – Vancea A, Bandici GE, Radovet D, Blidar CF, The acclimatization capacity of the *Saintpaulia* and of the *Chrysanthemum* exvitroplantlets, derived from vitroculture medium in which the saccharose was replaced with honey. *Analele Universității din Craiova*, XII (XLVIII), Secțiunea Biologie, 209-214, 2008.
- Purcărea C, Borbely MV, Salicylic acid involvement in wheat (*Triticum aestivum*) seedlings response to salt stress. *Scientific Papers, UASVM Bucharest, Series A*, LIV, 451-456, 2011.
- Sand C, Cachiță CD, Studiul comportamentului *in vitro* a unor explante de sfeclă de zahăr (*Beta vulgaris* L) prelevate de la familii și linii rezultate din procesul de ameliorare. In: Cachiță CD, Ardelean A (eds.), *Lucrările celui de-al XII -lea Simpozion Național de Culturi de Țesuturi și Celule Vegetale*, Jibou, intitulat „Fiziopatologia celulei vegetale în regim de vitrocultură”, Editura Daya Satu – Mare, 147 – 155, 1999.
- Sand C, Cachiță CD, Influența mediului de cultură asupra regenerării de plante din calus de sfeclă de zahăr (*Beta vulgaris* L var. *Sacharifera*). In: Cachiță CD, Ardelean A (eds.), *Lucrările celui de-al IX-lea Simpozion Național de Biotehnologie Vegetală*, Editura Universității “Ovidius” din Constanța, 151-158, 2000.
- Vancea A, Cachiță CD, Pătru DM, Adapted capacity of *Cymbidium hybridum* plantlets at *ex vitro* conditions, which was provided from culture medium with honey as sucrose replaced. *Analele Universității din Oradea – Fascicula Biologie*, IX, 213 – 218, 2002.